Research Note

Ten Records of Cercosporoid Fungi from the Lienhuachih Forest in Taiwan

Hsin-Hui Shih¹⁾ Yen-Tzu Fann²⁾ Jie-Hao Ou²⁾ Pei-Hsuan Lee¹⁾ Yung-Chun Chou¹⁾ Chi-Yu Chen^{2,3)}

[Summary]

Ten cercosporoid fungi were found in the Lienhuachih forest, Yuchi, Nantou County, westcentral Taiwan. They are *Pseudocercospora borreriae*, *P. eupatorii-formosani*, *P. brachypus*, *P. wedeliae*, *P. puerariicola*, *Cercospora bidentis*, *C. stachytarphetae*, *C. ageraticola*, *C. camptothecae*, and *Pseudocercosporella oxalidis*. Among them, *P. borreriae* and *P. brachypus* are first reported herein in Taiwan. Descriptions and illustrations are provided for these 2 species. **Key words:** cercosporoid fungi, new record, Lienhuachih.

Shih HH, Fann YT, Ou JH, Lee PH, Chou YC, Chen CY. 2022. Ten records of Cercosporoid fungi from the Lienhuachih forest in Taiwan. Taiwan J For Sci 37(2):157-65.

¹⁾Lienhuachih Research Center, Taiwan Forestry Research Institute, 43 Hualong Lane, Yuchih Township, Nantou 55543, Taiwan. 行政院農業委員會林業試驗所蓮華池研究中心,55543南投縣魚池 鄉五城村華龍巷43號。

²⁾ Department of Plant Pathology, National Chung Hsing Univ, 145 Xingda Rd., South Dist., Taichung 40227, Taiwan. 國立中興大學植物病理學系, 40227台中市南區興大路145號。

³⁾Corresponding author, e-mail:chiyu86@dragon.nchu.edu.tw 通訊作者。 Received April 2022, Accepted May 2022. 2022年4月送審 2022年5月通過。

研究簡報

臺灣蓮華池森林十種尾孢菌類真菌之紀錄

施欣慧¹⁾ 范晏滋²⁾ 歐玠皜²⁾ 李沛軒¹⁾ 周詠鈞¹⁾⁾ 陳啟予^{2,3)} 摘 要

於臺灣南投縣魚池鄉蓮華池地區採集到10種尾孢菌類真菌,分別為:豐花草假尾孢菌 (Pseudocercospora borreriae)、香澤蘭假尾孢菌(P. eupatorii-formosani)、細窄假尾孢菌(P. brachypus)、蟛蜞菊假尾孢菌(P. wedeliae)、葛藤假尾孢菌(P. puerariicola)、鬼針草尾孢菌(Cercospora bidentis)、長穗木尾孢菌(C. stachytarphetae)、藿香薊尾孢菌(C. ageraticola)、喜樹尾孢菌(C. camptothecae)和酢漿草假小尾孢菌(Pseudocercosporella oxalidis)。其中,豐花草假尾孢菌和細窄假尾 孢菌為臺灣新紀錄種。本文亦將對此新紀錄種進行詳細描述。

關鍵詞:尾孢菌類真菌、新紀錄、蓮華池。

施欣慧、范晏滋、歐玠皜、李沛軒、周詠鈞、陳啟予。2022。臺灣蓮華池森林十種尾孢菌類真菌之紀 錄。台灣林業科學37(2):157-65。

INTRODUCTION

Cercosporoid fungi are hyphomycetes resembling Cercospora that taxonomically belong to the family Mycosphaerellaceae (Ascomycota) (Crous and Braun 2003). Cercosporoid fungi are comprised of the genera Cercospora, Pseudocercospora, Ramularia, Passalora, Stenella, Cercosporella, Pseudocercosporella, etc., with the former 4 genera being the most prevalent. The major characters used for identification are the shape and pigmentation of the conidiophores and conidia, the texture of the conidial wall, and the presence or absence of conidial scars. Most cercosporoid fungi are plant pathogens causing leaf spots on ferns, gymnosperms, monocots, and dicots, and have high host specificity. Some of them are economically important pathogens of crops and trees, such as angular leaf spot of beans (Pseudocercospora griseola), leaf spot of sugar beets (Cercospora beticola), angular leaf spot of camptotheca trees (Cercospora camptothecae), mycosphaerella leaf disease of eucalypt trees (Crous, 1998), etc.

The earliest studies of cercosporoid fungi in Taiwan were conducted by Kaneyoshi Sawada (Sawada 1919, 1922, 1928, 1931, 1933, 1942, 1943a, 1943b, 1944, 1959), followed by Drs. Hsieh and Goh, who included nearly 240 species of cercosporoid fungi in their valuable book Cercospora and similar fungi from Taiwan (Hsieh and Goh 1990). In recent years, some new species and newly recorded species of cercosporoid fungi have continuously been published (Kirschner et al. 2004, Kirschner and Chen 2007, Kirschner and Okuda 2013, Kirschner 2014, Kirschner and Liu 2014, Kirschner and Wang 2015, Chen et al. 2020). To the present, approximately 410 species of cercosporoid fungi have been recorded in Taiwan. The Lienhuachih forest, known for its great biodiversity, is located at Yuchi Township, Nantou County, west-central Taiwan, at an elevation of 576~925 m. The annual average temperature is about 21°C, and annual average rainfall is about 2200 mm in the area. In a survey of cercosporoid fungi in the Lienhuachih forest, 10 species were obtained. Identification of these species was mainly based on host specificity, disease symptoms, and microscopic fungal characters. Two species are new records to Taiwan. Fresh cultures of these 2 species were obtained for subsequent internal transcribed spacer (ITS) sequencing. Thereafter, the identification was further justified.

MATERIALS AND METHODS

All fresh samples with disease lesions were collected by H. H. Shih at Lienhuachih, Nantou County, Taiwan. Samples were examined for the presence of cercosporoid fungi under a Leica MZ125 stereo microscope (Germany). To identify the fungi, detailed fungal microscopic characters were determined using a Leica DM2500 light microscope with an oil-immersion lens $(1000\times)$, and about 20~30 conidia and conidiophores were measured. Single spores were isolated for fungi first reported in Taiwan. Potato dextrose agar (PDA) medium was used to incubate the fungi at room temperature. Purified cultures were deposited at the Bioresource Collection and Research Center (BCRC; Hsinchu, Taiwan), and dry specimens were deposited at the Lienhuachih Research Center, Taiwan Forestry Research Institute (TFRILHC).

DNA was extracted as described by the protocol of Sambrook and Russell (2001). For DNA barcoding, primer pairs of ITS5 (5'-GGAAGTAAAAGTCGTAACAAGG-3')/ ITS4 (5'-TCCTCCGCTTATTGATATGC-3') (White et al. 1990) and V9G (5'-TTAC-GTCCCTGCCCTTTGTA-3') (Gerrits van den Ende and de Hoog 1999)/LR1 (5'-GGTTG-GTTTCTTTTCCT-3') (Vilgalys and Hester 1990) were used to amplify the internal transcribed spacer (ITS). A polymerase chain reaction (PCR) was performed with initial denaturation at 94°C for 5 min, followed by 35 cycles of 94°C for 30 s, 30 s at 55°C, and 60 s at 72°C, with a final extension of 10 min at 72°C. ITS sequences were sequenced using the same primers for the PCR amplification which was performed on an ABI PRISM 377 DNA sequencer at the Biotechnology Center of National Chung Hsing Univ (Taichung, Taiwan). Sequences were submitted through the submission system of the DNA Data Bank of Japan (DDBJ) (Mashima et al. 2016).

RESULTS

Ten cercosporoid fungi including 4 Cercospora species, 5 Pseudocercospora species, and 1 Pseudocercosporella species were collected and studied during 2020~2022 in the Lienhuachih forest. They are Cercospora ageraticola on Ageratum houstonianum (Asteraceae), C. bidentis on Bidens pilosa (Asteraceae), C. camptothecae on Camtoptheca acuminate (Nyssaceae), C. stachytarphetae on Stachytarpheta jamaicensis (Verbenaceae), Pseudocercospora brachypus on Ampelopsis cantoniensis (Vitaceae), P. eupatorii-formosani on Chromolaena odoratum (Asteraceae), P. puerariicola on Pueraria montana var. lobata (Fabaceae), P. borreriae on Spermacoce latifolia (Rubiaceae), P. wedeliae on Wedelia triloba (Asteraceae), and Pseudocercosporella oxalidis on Oxalis debilis var. corymbosa (Oxalidaceae). Among them, P. borreriae and P. brachypus were newly recorded in Taiwan. The collection information with morphological descriptions for the 2 new Taiwan records are as follows.

Cercospora ageraticola Goh & Hsieh, Trans Mycol Soc Rep China 4(2-3):40, 1989 (藿香 薊尾孢菌)

Specimens examined: On leaves of Ag-

eratum houstonianum (Asteraceae), 30 Nov. 2021 (TFRILHC-1101130-7); 11 Feb. 2022 (TFRILHC-1110211-3).

Early report in Taiwan: Hsieh and Goh (1990).

Cercospora bidentis Tharp, Mycologia 9(2):108, 1917 (鬼針草尾孢菌) = Cercospora bidentis Marchal & Steyaert, Bull Soc Roy Bot Belgique 61:167, 1929. = Cercospora bidentis-pilosae Sawada, Taiwan Agric Res Inst Rep 85:98, 1943.

Specimens examined: On leaves of *Bidens pilosa* (Asteraceae), 5 Nov. 2021 (TFRILHC-1101105-9); 25 Nov. 2021 (TFRILHC-1101125-1).

Early report in Taiwan: Hsieh and Goh (1990).

Cercospora camptothecae Tai, Lloydia 11:39, 1948 (喜樹尾孢菌)

Specimen examined: On leaves of *Camtoptheca acuminate* (Nyssaceae), 2 Dec. 2021 (TFRILHC-1101202-4).

Early report in Taiwan: Chen (1968).

Cercospora stachytarphetae Ellis & Everh., Missouri Bot Gard Ann Rep 9:120, 1898 (長 穗木尾孢菌)

Specimens examined: On leaves of *Stachytarpheta jamaicensis* (Verbenaceae), 15 Jan. 2021 (TFRILHC-1100115-1); 17 Nov. 2021 (TFRILHC-1101117-1).

Early report in Taiwan: Hsieh and Goh (1987).

Pseudocercospora borreriae (Ellis & Everh.) Deighton, Mycol Pap 140:140, 1976. Fig. 1. (豐花草假尾孢菌)

≡ Cercospora borreriae Ellis & Everh. Proc Acad Nat Sci Phil 46(3):379, 1894

Leaf spots amphigenous, irregular to circular, 5~15 mm in diam., pale-brown to

gravish-brown in center, with dark-brown border, sometimes many small leaf spots fusing to appear like leaf blight. Stromata amphigenous, well-developed, immersed. Conidiophores densely fasciculate, cylindrical, straight to slightly curved, pale-brown, unbranched or rarely branched, conically truncate at apex, $76 \sim 165 \times (2.5 \sim) 3 \sim 5 \mu m$ with $3 \sim 7$ septa, conidial scars not thickened. Conidia solitary, straight or slightly curved, subhyaline to pale-olivaceous, cylindrical to obclavate or cylindric-obclavate, $45 \sim 105 \times$ $3.7 \sim 5.5 \,\mu\text{m}$ with $3 \sim 7(\sim 8)$ septa, with round to obtuse apex and conically truncated base, 2~3 µm wide, hila neither thickened nor darkened. Colony on PDA slow-growing, dome-shaped, lobate, white, gray, to dark-gray.

Specimens examined: On leaves of *Spermacoce latifolia* (Rubiaceae), 23 Oct. 2020 (TFRILHC-1091023-1; living culture: BCRC FU31708); 23 Aug. 2021 (TFRILHC-1100823-1); 23 Sept. 2021 (TFRILHC-1100923-2); 19 Oct. 2021 (TFRILHC-1101019-3); 5 Nov. 2021 (TFRILHC-1101105-5).

Note: Pseudocercospora borreriae, based on Cercospora borreriae, was originally reported on *Borreria micrantha* (= Spermacoce remota) (Ellis and Everhart 1894), and has now been widely reported on many other plant genera in Borreria, Mitracarpus, and Spermacoce (Farr and Rossman 2022), all belonging to the family Rubiaceae. The original description (Ellis and Everhart 1894) of this species was too simple to tell characteristic differences from other species. Nevertheless, the justification for this species was largely based on host identity, i.e., Spermacoce affinity. The fungus isolated from Spermacoce latifolia in Taiwan conforms to the description of Pseudocercospora borreriae by Dennis (1970) on Spermacoce latifolia, and Guo (2012) on Borreria stricta. The ITS sequence

160

(accession no. LC700328) of 496 base pairs was successfully obtained in this study. Unfortunately, there was no sequence named under Pseudocercospora borreriae in GenBank for comparison. In a blast search, the highest hit was NR147312.1, namely Pseudocercospora richardsoniicola, with 99.17% similarity. Pseudocercospora richardsoniicola was reported on Richardsonia sp. and Richardia brasiliensis (Rubiaceae) (Silva et al. 2016), which are phylogenetically closely related to Spermacoce (Groeninckx et al. 2009). Pseudocercospora richardsoniicola is morphologically indistinguishable from *P. borreriae* based on the available description. They are probably the same fungus, but type sequences of both species must be compared. However, according to the host identity, Pseudocercospora borreriae was applied to name the Taiwan material.

Pseudocercospora coremioides was reported on *Diodia*, a genus phylogenetically closely related to *Spermacoce* (Groeninckx et al. 2009). However, *P. coremioides* can be distinguished by having longer and solitary conidiophores (40~300 μ m) bearing wider conidia (4~6.5 μ m) with more septa (3~12) (Braun and Urtiaga 2013).

Pseudocercospora brachypus (Ellis & Everh.) Liu & Guo, Acta Mycol Sin 11:128, 1922; The Genus *Pseudocercospora* in China, p 352, 1995. Fig. 2. (細窄假尾孢菌)

≡ Cercospora brachypus Ellis & Everh., J Mycol 8:71, 1902.

Leaf spots angular to irregular, $4\sim10$ mm in diam., brown, dark-reddish-brown to darkbrown with a blackish-brown margin. Secondary mycelium well-developed, hyphae subhyaline to pale-olivaceous, branched, septate, arcuate, 1.5~2.5 µm wide, bearing secondary conidiophores. Stromata nearly globular, dark-brown, 20~45 µm wide. Conidiophores densely fasciculate, pale-brown, cylindric, irregular in width, mostly straight and erect, unbranched, $0(\sim 1)$ septate, obtuse at apex, $10\sim 25 \times 2.3\sim 3.6 \mu m$, conidial scars not thickened. Conidia subhyaline to pale-olivaceous, cylindric, straight to slightly curved, $(2\sim)3\sim 10$ septate, subobtuse to rounded at apex, obconically truncate at base, $23\sim 68(\sim 85) \times 1.9\sim 3.7$ μm , hila neither thickened nor darkened, $1\sim 1.5 \mu m$ wide. Colony on PDA slow-growing, dome-shaped, lobate, gray at periphery, dark-gray to dark-olivaceous-gray in center.

Specimens examined: On leaves of *Ampelopsis cantoniensis* (Vitaceae), 24 Nov. 2021 (TFRILHC-1101124-1; living culture: BCRC FU31709); 30 Nov. 2021 (TFRIL-HC-1101130-2); 20 Jan. 2022 (TFRIL-HC-1110120-2).

Note: Two Pseudocercospora species, P. brachypus and P. ampelopsis, have been reported on species of the host genus Ampelopsis (Guo and Hsieh 1995, Crous et al. 2013). In addition to Ampelopsis brevipedunculata (Vitaceae), P. brachypus was also reported on Parthenocissus tricuspidata (Vitaceae) and Vitis balanserma (Vitaceae), while P. ampelopsis was only reported on Ampelopsis glandulosa var. heterophylla (Vitaceae). The fungus of the Taiwan material conforms to the description of P. brachypus by Guo and Hsieh (1995) and significantly differs from P. ampelopsis in the morphology of the conidiophores and conidia. Pseudocercospora ampelopsis is distinct in having larger conidiophores (20~80 × $(2.5\sim)3\sim5(\sim6)$ µm) with more septa $(3\sim6)$ bearing larger conidia $((35\sim)40\sim90(\sim110))$ × $3\sim5(\sim6)$ µm) (Crous et al. 2013). Unfortunately, there was no sequence named under Pseudocercospora brachypus in GenBank for comparison. In a blast search using the ITS sequence (accession no. LC700327) of the Taiwan material, the high-

est hits were sequences under the names of P. ampelopsis (DQ303088) (Crous et al. 2013) and P. flavomarginata (MH863876/ GU269799) reported on Eucalyptus spp. (Myrtaceae), P. struthanthi (NR147304/ KT290141) reported on Struthanthus flexicaulis (Loranthaceae), and P. schizolobii (GQ852765/GQ885903) reported on Eucalyptus camaldulensis and Schixolobium parahyba (Fabaceae). Pseudocercospora flavomarginata is distinct in having larger stromata (56 \times 47 μ m), larger conidiophores $((18\sim)32\sim36(\sim53)\times(2\sim)3\sim4(\sim5) \mu m)$, larger conidia ((28~)46~54(~90) × (2~)3(~4) μ m), and different numbers of septa of conidiophores and conidia (Hunter et al. 2006); P. struthanthi in having wider conidiophores $(7.5 \sim 31 \times 3 \sim 5.5 \ \mu m)$, larger conidia (41 ~ 83.5 \times 3~4 µm), and more septa (0~3) of the conidiophores (Silva et al. 2016); P. schizolobii in having larger conidiophores $(30 \sim 70 \times 10^{-70})$ $3.5 \sim 4 \mu m$) and conidia ((22~) 40~60 (~90) \times (3~) 3.5 (~4) µm) and more septa (1~4) of the conidiophores (Crous et al. 2009). Pseu*docercospora brachypus* is the name best accommodating the Taiwan material.

Pseudocercospora eupatorii-formosani Yen ex Guo & Hsieh, Mycosystema Monographicum 2:67, 1995 (香澤蘭假尾孢菌)

 \equiv Cercospora eupatori-formosani Sawada, Taiwan Agric Res Inst Rep 86:169, 1943.

Specimen examined: On leaves of *Chromolaena odoratum* (Asteraceae), 5 Nov. 2021 (TFRILHC-1101105-3).

Early report in Taiwan: Hsieh and Goh (1990).

Pseudocercospora puerariicola (Yamam.) Deighton, Mycol Pap 140:151, 1976 (葛藤假 尾孢菌)

≡ Cercospora puerariicola Yamamoto, Trans Sapporo Nat Hist Soc 13:142, 1934.

Specimens examined: On leaves of *Pueraria montana var. lobata* (Fabaceae), 24 Sept. 2021 (TFRILHC-1100924-1); 18 Oct. 2021 (TFRILHC-1101018-2); 17 Feb. 2022 (TFRILHC-1110217-3).

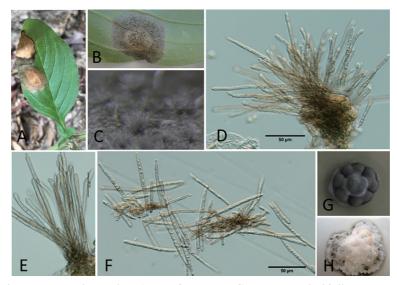


Fig. 1. *Pseudocercospora borreriae*. A, Leaf spots on *Spermacoce latifolia*; B, close-up of lesion; C, close-up of leaf spot with superficial sporulation structures; D and E, fasciculate conidiophores; F, conidia; G and H, colony on PDA.

Early report in Taiwan: Hsieh and Goh (1990).

Pseudocercospora wedeliae (Kar & Mandal) Deighton, Mycol Pap 140:155, 1976 (蟛蜞菊 假尾孢菌)

■ Cercospora wedeliae A.K. Kar &
M. Mandal, Trans Br Mycol Soc 54(3):428, 1970.

Specimens examined: On leaves of *Wedelia triloba* (Asteraceae), 24 Nov. 2021 (TFRILHC-1111124-2); 25 Jan. 2022 (TFRILHC-1111124-2-1).

Early report in Taiwan: Kirschner (2014).

Pseudocercosporella oxalidis (Goh & Hsieh) U. Braun, Nova Hedwigia 55(1-2):218, 1992 (酢漿草假小尾孢菌)

Specimens examined: On leaves of *Oxalis debilis* var. *corymbosa* (Oxalidaceae), 30 Nov. 2021 (TFRILHC-1101130-4); 8 Dec. 2021 (TFRILHC-1101208-1); 3 Mar. 2022

(TFRILHC-1110303-1).

Early report in Taiwan: Goh and Hsieh (1989).

LITERATURE CITED

Braun U, Urtiaga R. 2013. New species and new records of cercosporoid hyphomycetes from Cuba and Venezuela (Part 2). Mycosphere 4(2):174-214.

Chen CC. 1968. Survey of epidemic diseases of forest trees in Taiwan V. Plant Prot Bull (Taiwan, R.O.C.) 10(4):13-5.

Chen CH, Hsieh SY, Yeh YH, Kirschner R. 2020. *Cladocillium musae*, a new genus and species of cercosporoid fungi (Mycosphaerellaceae) on wild banana in Taiwan. Mycol Progr 19(9):837-43.

Crous PW. 1998. *Mycosphaerella* spp. and their anamorphs associated with leaf spot diseases of *Eucalyptus*. Mycol Mem 21:1-170.

Crous PW, Braun U. 2003. *Mycosphaerella* and its anamorphs: 1. Names published in

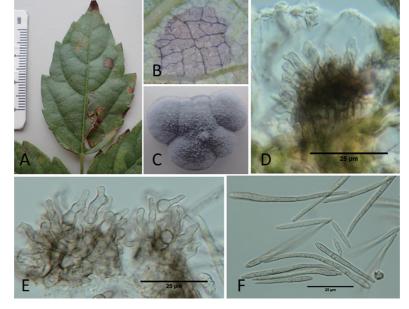


Fig. 2. *Pseudocercospora brachypus*. A, Leaf spots on *Ampelopsis cantoniensis*; B, close-up of lesion; C, colony on PDA; D and E, fasciculate conidiophores; F, conidia.

Cercospora and *Passalora*. CBS Biodiversity Series 1. 571 p.

Crous PW, Braun U, Hunter GC. 2013. Phylogenetic lineages in *Pseudocercospora*. Stud Mycol 75:37-114.

Crous PW, Summerbell RC, Carnegie AJ, Wingfield MJ, Hunter GC, Burgess TI, et al. 2009. Unravelling *Mycosphaerella*: Do you believe in genera? Persoonia 23:99-118.

Deighton FC. 1976. Studies on *Cercospora* and allied genera. VI. *Pseudocercospora* speg., *Pantospora* cif. and *Cercoseptoria* petr. Mycol Pap 140:140.

Dennis RWG. 1970. Fungi of Venezuela and adjacent countries. Kew Bull Add Seri 3:1-531.

Ellis JB, Everhart BM. 1894. New species of fungi from various localities. Proc Acad Nat Sci Phil 46(3):379.

Farr DF, Rossman AY. Fungal databases, US National Fungus Collections, ARS, USDA. Available at https://nt.ars-grin.gov/fungaldatabases/. Accessed March 8, 2022.

Gerrits van den Ende AHG, de Hoog GS. 1999. Variability and molecular diagnostics of the neurotropic species *Cladophialophora bantiana*. Stud Mycol 43:151-62.

Groeninckx I, Dessein S, Ochoterena H, Persson C, Motley TJ, Kårehed J, et al. 2009. Phylogeny of the herbaceous tribe Spermacoceae (Rubiaceae) based on plastid DNA data 1. Ann Missouri Bot Gard 96:109-32.

Guo YL. 2012. Studies on *Cercospora* and allied genera of China XVI. Mycosystema 31(4):476-9.

Guo YL, Hsieh WH. 1989. New species of *Cercospora* and allied genera of Taiwan. Bot Bull Acad Sin 30(2):117-32.

Guo YL, Hsieh WH. 1995. The genus *Pseudocercospora* in China. Beijing, China: International Academic Publishers. 388 p.

Hsieh WH, Goh TK. 1987. Studies on Cercospora and allied genera of Taiwan (I). Trans Mycol Soc ROC 2(1):23-36.

Hsieh WH, Goh TK. 1990. *Cercospora* and similar fungi from Taiwan. Taipei, Taiwan: Maw Chang Book Co. 376 p.

Hunter GC, Crous PW, Wingfield BD, Pongpanich K, Wingfield MJ. 2006. Pseudocercospora flavomarginata sp. nov., from Eucalyptus leaves in Thailand. Fungal Divers 22:71-90.

Kirschner R. 2014. A new species and new records of cercosporoid fungi from ornamental plants in Taiwan. Mycol Progr 13(3):483-91.

Kirschner R, Chen CJ. 2007. Foliicolous hyphomycetes from Taiwan. Fungal Divers 26:219-39.

Kirschner R, Liu LC. 2014. Mycosphaerellaceous fungi and new species of *Venustosynnema* and *Zasmidium* on ferns and fern allies in Taiwan. Phytotaxa 176(1):309-23.

Kirschner R, Okuda T. 2013. A new species of *Pseudocercospora* and new record of *Bartheletia paradoxa* on leaves of *Ginkgo biloba*. Mycol Progr 12:421-6.

Kirschner R. Piepenbring M, Chen CJ. 2004. Some cercosporoid hyphomycetes from Taiwan, including a new species of *Stenella* and new reports of *Distocercospora pachyderma* and *Phacellium paspali*. Fungal Divers 17:57-68.

Kirschner R, Wang H. 2015. New species and records of mycosphaerellaceous fungi from living fern leaves in East Asia. Mycol Progr 14:65.

Mashima J, Kodama Y, Kosuge T, Fujisawa T, Katayama T, Nagasaki H, et al. 2016. DNA data bank of Japan (DDBJ) progress report. Nucleic Acids Res 44:D51-7.

Sambrook J, Russell RW. 2001. Molecular cloning: a laboratory manual, 3rd ed. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.

Sawada K. 1919. Descriptive catalogue of the Formosan fungi I. Taiwan Agric Exp Stat Special Bull 19:666-84.

164

Sawada K. 1922. Descriptive catalogue of the Formosan fungi II. Dept Agric Govern Res Inst Taiwan Rep 2:139-64.

Sawada K. 1928. Descriptive catalogue of the Formosan fungi IV. Dept Agric Govern Res Inst Taiwan Rep 35:105-13.

Sawada K. 1931. Descriptive catalogue of the Formosan fungi V. Dept Agric Govern Res Inst Taiwan Rep 51:126-31.

Sawada K. 1933. Descriptive catalogue of the Formosan fungi VI. Dept Agric Govern Res Inst Taiwan Rep 61:94-6.

Sawada, K. 1942. Descriptive catalogue of the Formosan fungi VII. Dept Agric Govern Res Inst Taiwan Rep 83:159-69.

Sawada K. 1943a. Descriptive catalogue of the Formosan fungi VIII. Dept Agric Govern Res Inst Taiwan Rep 85:98-126.

Sawada K. 1943b. Descriptive catalogue of the Formosan fungi IX. Dept Agric Govern Res Inst Taiwan Rep 86:165-74.

Sawada K. 1944. Descriptive catalogue of the Formosan fungi X. Dept Agric Govern Res Inst Taiwan Rep 87:79-90.

Sawada K. 1959. Descriptive catalogue of the Taiwan (Formosan) fungi XI. Special Publ Coll Agric Natl Taiwan Univ. 8:211-77.

Silva M, Barreto RW, Pereira OL, Freitas NM, Groenewald JZ, Crous PW. 2016. Exploring fungal mega-diversity: *Pseudocercospora* from Brazil. Persoonia 37:142-72.

Vilgalys R, Hester M. 1990. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. J Bacteriol 172:4238-46.

White TJ, Bruns T, Lee S, Taylor J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis N, Gelfand D, Sninsky J, White T, editors. PCR protocols: a guide to methods and applications. New York: Academic Press. p 315-22.